SPECIFICATION

Please replace paragraph [0045] on pages 15 and 16 with the following amended paragraph [0045].

[0045] MTT cell proliferation assay: The cells were counted and transferred to 96 well microtiter plates, and incubated for 24 h prior to the addition of test compounds. The cell numbers used for each cancer cell lines were 6000, 3000, 4000, 3000 and 5000 per well for SF-268, NCI H460, MCF-7, HCT-116, and AGS, respectively. Test compounds were dissolved in DSO and diluted with sterile RPMI-1640 media as necessary to obtain the appropriate concentration. The test solutions were then added to the wells containing cells in 100-µL aliquots to obtain final appropriate concentrations. The final concentration of DMSO in each well was 0.2%. Test compounds, positive control, and blank control (DMSO in media) were incubated with all five cell-lines for 48 h, after which MTT solution (5 mg/mL in PBS solution) was added into each well in 25-µL aliquots. The plates then were wrapped in aluminum foil and incubated for three hours at 37°C with 5% CO_2 and 80% RH. The RPMI media, MTT

and floating cells from each well were removed and aliquots of DMSO (20 µL) added into each sample well to dissolve the purple formazan crystals. The plates were then shaken for eight minutes on a gyrorotary shaker after which the absorbence of the contents of each well was recorded with an automated microplate reader (model EL800, Bio-Tek Instruments, Inc., Winooski, VT) at 570 nm. The experiments were performed in triplicate at concentrations of 25, 50, 100 and 200 µg/mL. Cell viability was determined by comparing the average absorbance of three test wells verses that of the blank control wells. Results are expressed in a line graph as the percentage of cell viability against concentration of compounds in Figures ~~6 to 10~~ 5 to 7.